THE BIOSYNTHESIS OF HAZIMICINS: POSSIBLE ORIGIN OF ISONITRILE CARBON

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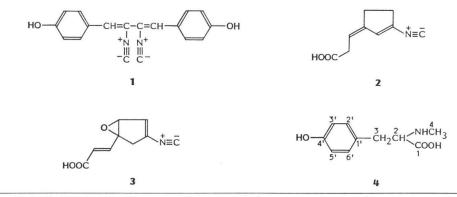
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Hazimicin factors 5 (R, R+S, S) and 6 (R, S), having comparable physico-chemical properties, are members of a new class of broad-spectrum antibacterial and antifungal antibiotics recently isolated from Micromonospora echinospora var. challisensis SCC 1411. The antibiotics are active against various Gram-positive, Gram-negative bacteria and yeasts, e.g., Staphylococcus aureus, Micrococcus luteus, Bacillus subtilis, Escherichia coli, Pseudomonas aeruginosa and Candida albicans. The details of taxonomy, biological properties¹⁾ and the structures²⁾ have been reported from our laboratories. The presence of an isonitrile $(-N^+ \equiv C^-)$ function was quite unusual and its biogenetic origin has been intriguing eversince the isolation of xanthocillin (1), the Pencillium metabolite³⁾. Carbon-14 labeling experiments indicated the incorporation of tyrosine, but not of methionine or formate into 1^{4} . More recently the biosyntheses of isonitrile containing antibiotics, 2 and 3, isolated from Trichoderma hamatum (Bon.) Bain. aggr. were reported to originate from tyrosine, however, the origin of the

isonitrile carbon was not $addressed^{5,6}$. The lack of data on the origin of an isonitrile carbon prompts us to report on the biosynthesis of hazimicins.

The fermentation was carried out either in flasks or 14-liter tanks according to the published procedure¹⁾. First the data on the incorporation of ¹⁴C-labeled precursors (conc 0.20 μ Ci/ml), Na-[1-¹⁴C]acetate, Na-[2-¹⁴C]acetate, Na-[3-¹⁴C]-pyruvate, L-[methyl-¹⁴C]methionine, L-[1-¹⁴C]-tyrosine, DL-[3-¹⁴C]tyrosine, [1-¹⁴C]formaldehyde were obtained. Then the ¹³C-labeled DL-[3-¹³C]-tyrosine (25 mg/liter), L-[methyl-¹³C]methionine (200 mg/liter) and DL-[*N*-methyl-¹³C]tyrosine* (4, conc 25 mg/liter) were added individually to the culture at the optimal time; additions were done at either day two or three.

The fermentation broth was extracted twice with double volumes of EtOAc and the combined extract was further concentrated. For 14Clabeled studies, this extract was chromatographed in chloroform - methanol (9:1) on LK6DF Whatman silica plates and scanned for radioactivity. Duplicate plates were tested by bioautography on Micrococcus luteus plates. For ¹³C NMR, the EtOAc extract was purified on Sephadex LH-20 column using ethanol as a solvent. Since the incorporations of ¹⁴C-labeled precursors in components 5 and 6 were comparable, it was not necessary to separate them for the present study. Fully decoupled ¹³C NMR spectra of the natural abundance ¹³C and the enriched antibiotic samples were recorded in DMSO- d_{θ} on a Varian XL-100-15 NMR Spectrometer operating at 25.2 MHz. Percentage enrichments obtained from both components of hazimicin are presented in Table 1.



* $DL-[N-Methyl-^{13}C]$ tyrosine was prepared by N-methylation using $^{13}CH_3I$ of N-trifluoroacetyl benzyloxy tyrosine and subsequent removal of the protecting groups.

Table 1. Incorporation of ¹⁴C- and ¹³C-labeled percursors.^a

C^b	Hazimicin (ppm) Factors 5 & 6	[3- ¹⁸ C]- Tyrosine		L-[<i>Methyl</i> - ¹³ C]- methionine	
		I/Io	[% ¹⁴ C]	I/Io	[% ¹⁴ C]
1	167.0	1.00		1.00	
2	58.5	1.27		1.11	
3	37.7	8.11	14~16	0.97	
1'	126.0	1.12		1.05	
2'	129.0	1.14		0.92	
3'	125.7	1.32		1.13	
4'	153.5	1.00		1.00	
5'	115.6	1.13		1.09	
6'	132.1	0.80		0.78	
4	158.1			1.45	1.5

- ^a Ratio of normalized intensities of the same carbon for enriched (I) and natural abundance (Io) samples. The spectra were obtained under essentially identical conditions with s/n of 37:1 for the methionine experiment.
- ^b The chemical shifts of the monomer are: δ (ppm) C-1=167.0, C-2=58.7, C-3=37.6, C-1'=125.9, C-2',6'=130.2, C-3',6'=115.0, C-4'=156.4 and C-4=158.0.

The ¹⁴C-labeling studies indicated that only DL-tyrosine (16%), [2-14C]acetate (0.04%) and methionine were incorporated into the antibiotic complex. In the case of methionine, however, 1.5% of radioactivity was found in components 5 and 6, the rest (3.5%) ended up in the bioinactive components. The addition of DL-[3-13C]tyrosine (25 mg/liter) highly enriched C-3 with percentage comparable to the enrichment of $14 \sim 16\%$ obtained from 14C-labeled tyrosine. The incorporation of [13C]methionine at the above concentration was considerably lower; the intensity of C-4 enhanced by $15 \sim 20$ %. When 8 times the initial concentration was used a significant enrichment of C-4 as well as a $3 \sim 4$ fold increase in the production of hazimicins 5 and 6 was observed. It appears, therefore, that isonitrile carbon originated from L-[methyl-13C]methionine, and it is probable that -NHCH₃ is converted to -NC through the intermediacy of -NHCHO based upon chemical analogy^{2,7)}.

$$-HNCH_{3} \rightarrow [-NH-CH_{2}OH] \rightarrow -HNCHO$$
$$\rightarrow -N=C=O \rightarrow -N^{+} \equiv C^{-}$$

The above sequence was further supported by the formation of the *N*-formyl product in some fermentations (-CHNH-CHO, δ_{CH} =4.9 became a triplet on D₂O exchange of δ_{NH} =8.82, δ_{CHO} =

Fig. 1. Biosynthesis of hazimicins.



8.02s)[†]. However attempts to incorporate ¹⁴C-labeled formaldehyde into hazimicins failed as with the *Penicillium* metabolite⁴⁾. Similarly the addition of DL-[*N*-methyl-¹³C]tyrosine failed to show any incorporation of C-4 at δ 158.1 indicating either lack of incorporation or demethylation in the initial stages of the fermentation. Thus hazimicins (Fig. 1) are biosynthesized from tyrosine and methionine; the latter established an origin of isonitrile carbon.

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[†] After the completion of our work, Prof. SCHEUER, et al., reported that in the sponge Hymeniacidon sp. an isolated isocyanoterpene coexisted with a formamide and an isothiocyanate and the -NC function is transformed to -NHCHO and -NCS (J. Am. Chem. Soc. $106:2447 \sim 2448, 1984$). It is conceivable, therefore, that the isocyano antibiotic was microbially converted to the formamide. 1229, 1981

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